

Fast color kit method technical information

Security card code 12-103

Product code 12-103

Pack 3x100 ml or on request.

Stability of product properly conserved at 15-20°C 24 months

Produce in Italy by

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En cas d'urgence, contactez votre unité de poison contre le plus proche

UE

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Suisse

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Application.

Fast color kit rapid staining set for microscopy" is used for human-medical cell diagnosis and serves the purpose of the haematological and cytological investigation of sample material of human origin. It is a ready-to-use staining kit that when used together with other in vitro diagnostic products from our portfolio makes target structures (by fixing, staining, where necessary counterstaining, mounting) in haematological and clinico-cytological specimen materials, for example smears of whole blood and bone marrow, evaluable for diagnostic purposes. Fast color kit is used for the rapid staining of haematological and cytological specimen material to produce a staining that corresponds to the Pappenheim stain, shortening the classic staining method to a staining time of 15-20 second

Principle.

Fast color is a fast-acting variation of May Grünwald Giemsa stain.

In hematology, it allows differential staining of blood smears (white blood cell count, morphological erythrocyte analysis and testing of parasites). It is ideal for emergencies' and for studying smears in specimens showing as positive on cell counter machine. In cytology, it allows cytological examinations of fluid and of fine-needle punctures. In pathology, it allows emergency examination of frozen tissue section completed during surgical operation. Stains present in this kit are the same utilized in traditional May Grünwald Giemsa. In this case, the rapidity with which the coloration process is completed is due to the fast dissociation of the tyazinic and others stain contained that they rented extremely rapid the absorption of cellular structures.

Blood smear

Dip the slides 5 times for 1 second into solution 1. Drain the surplus onto filter paper.

Dip the slides 5 times for 1 second into solution 2. Drain the surplus onto filter paper.

Dip the slides 5 times for 1 second into solution 3. Drain the surplus onto filter paper.

Briefly wash in running water to remove excess and dip into alcohol 5-6 time.

Malaria (Plasmodium)

Dip the slides 3 times for 1 second into solution 1. Drain the surplus onto filter paper.

Dip the slides 2 times for 1 second into solution 2. Drain the surplus onto filter paper.

Dip the slides 2 times for 1 second into solution 3. Drain the surplus onto filter paper.

Briefly wash in running water to remove excess and dip into alcohol 5-6 time.

Cytological of fluid and fine-needle punctures

Dip the slides 5 times for 1 second into solution 1. Drain the surplus onto filter paper.

Dip the slides 5 times for 1 second into solution 2. Drain the surplus onto filter paper.

Dip the slides 10 times for 1 second into solution 3. Drain the surplus onto filter paper.

Wash carefully in running water to remove excess and dip into alcohol 5-6 time.

Histological slides

Dip the slides 10 times for 1 second into solution 1. Drain the surplus onto filter paper.

Dip the slides 15 times for 1 second into solution 2. Drain the surplus onto filter paper.

Dip the slides 15 times for 1 second into solution 3. Drain the surplus onto filter paper.

Dehydrate and mount with DdMount.

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Pneumocystis carinii Cryptosporidia Leishmania Cryptococcus Microfilaria Toxoplasma

Dip the slides 10 times for 1 second into solution 1. Drain the surplus onto filter paper.

Dip the slides 10 times for 1 second into solution 2. Drain the surplus onto filter paper.

Dip the slides 10 times for 1 second into solution 3. Drain the surplus onto filter paper.

Briefly wash in running water to remove excess and dip into alcohol 5-6 time.

Frozen tissue section

Dip the slides 10 second in solution 1. Drain the surplus onto filter paper.

Dip the slides 15 second in solution 2. Drain the surplus onto filter paper.

Dip the slides 20 second in solution 3. Drain the surplus onto filter paper.

Dehydrate and mount with DdMount.

Blood cells result:

Nuclei:	chromatin:	purple
Leucocytes:	cytoplasm with out ARN:	light pink
	eosinophilic granules:	orange brown
	basophilic granules:	dark purplish blue
	neutrophilic granules:	purple ± intense
Lymphocytes:	cytoplasm with ARN:	blue
	cytoplasm with out ARN:	light blue
	azurophilic granules:	red
Monocytes:	cytoplasm:	grey-blue
Erythrocytes:		light red
Platelets:	chromomer:	purplish red
	hyalomere:	bluish
Blood parasites:	nucleus	red
(Malaria)	cytoplasm	blue

Please note

Staining time may change according to thickness, and dryness' of slides.

* Technical's note: staining time vary according to age, types of solutions, thickness of sections, et. When Gill (code 09-178) modified solution is used, get the best result, staining time (maximum 1-5 minutes), for best change in color, wash quickly in tap water, and then in Scott acidulated solution, (code 00-136). For sections fixed in Bouin, we recommend the use of haematoxylin modified acid AB (code 09-183). Please note the alcoholic loses eosin stain with the use, of the days are stretched over time colouring. If you are using purified eosin, check the time, and possibly diluted in ethyl alcohol 96°C, if the cytoplasmic staining was too strong. Before use, filter the following solutions; alcoholic eosin, eosin phloxine; Harris haematoxylin, Gill's haematoxylin. The acidified aqueous solution of eosin is prepared by slowly adding glacial acetic acid. Follow normal precautions for laboratory reagents. Dispose of waste according to regulations at the local, regional or national level. Refer to Data Sheet Material Safety Data for updated information on risks, hazards and safety associated with the use of these products.

* Risk and Safety Statements outside the EU.

The eosin solution in alcohol is flammable and harmful. Harmful by inhalation, in contact with skin or if swallowed. Harmful: possible risk of irreversible effects through inhalation, in contact with the skin or by ingestion. Irritating to eyes, respiratory system and skin. Keep away from sources of ignition - No smoking. Wear suitable protective clothing and gloves. In case of accident or if you feel unwell, seek medical attention immediately (show the label where possible). Target organs: eyes and nerves. Eosin in aqueous solution. Caution: substance not yet fully tested. Avoid contact and inhalation of the solution of Harris haematoxylin. Organs: heart and nerves. Solutions based hemallum are harmful. Harmful if swallowed. Irritating to eyes, respiratory system and skin. In case of contact with eyes, rinse immediately with plenty of water and seek medical attention. Wear suitable protective clothing. Organs affected: liver and kidneys. In case of accident or if you feel unwell, seek medical attention immediately (show the label where possible).

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*** Risk and Safety Statements (U.E.)**

The eosin solution in alcohol is highly flammable and harmful. Highly flammable. Harmful by inhalation, in contact with skin or if swallowed. Harmful: possible risk of irreversible effects through inhalation, in contact with the skin or by ingestion. Keep away from sources of ignition - No smoking. Wear suitable protective clothing and gloves. In case of accident or if you feel unwell, seek medical attention immediately (show the label where possible).

Eosin in aqueous solution. Caution: Substance not yet fully tested. Solution of hemallum. Do not breathe vapors. Avoid contact with skin and eyes. Gill haematoxylin Solutions are harmful. Harmful if swallowed. Irritating to eyes, respiratory system and skin. In case of contact with eyes, rinse immediately with plenty of water and seek medical attention. Wear suitable protective clothing.

*** Sample preparation**

All samples must be treated according to the technology. All samples must be marked so as to be easily identified. Tools should be used for sampling and sample preparation, which must be observed strictly to manufacturer's instructions about the application and instructions. Diagnostics. The diagnosis should be performed only by authorized and trained persons. Valid nomenclatures must be used. Further tests must be selected and implemented according to recognized methods. Instructions for use. To avoid errors, the staining process must be performed by qualified personnel. For professional use only. Must observe the National guidelines for work safety and quality assurance. Microscopes are used according to the standard. Protection against infection. Must be taken with laboratory guidelines for the protection against infection. Instructions for disposal. The solutions used and those have expired must be disposed of as special waste according to local regulations regarding disposal of waste.

Endnotes

1 The timing suggested in the leaflet are approximate and may vary according to your specific needs. If they are used intensively, for staining solutions may lose their dyes, so it is necessary to extend the time of staining solutions, or replace with new products.

2. Include positive control slides in each session.

3. Some hydraulic systems deliver acidic water, unsuitable for use for the part of the procedure for the blue coloration. If tap water is acidic, instead using a dilute alkaline solution, for example, water buffered by Scott.

4. The presence of purple or red-brown nuclei a blue color indicates unsatisfactory.

5. If you over-eosin staining, nuclear staining may be masked. If done correctly, with eosin staining shows a three-tone effect. To increase the differentiation of eosin, extend the time of immersion in alcohol, or use a first alcohol with a higher water content. You can adjust the times of immersion in alcohol to obtain an adequate eosin staining.

6. We do not recommend the addition of stock solution in the working solutions of haematoxylin and eosin.

7. Avoid excessive drag (carryover) of water solutions in alcoholic eosin.

8. The data generated by this procedure are to be used only to support the diagnosis and should be evaluated in conjunction with other tests and diagnostic data

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