

Technical Data Sheet

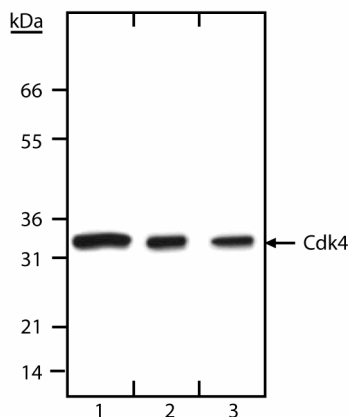
Purified Anti-Human Cdk4 (N-Term)

Product Information

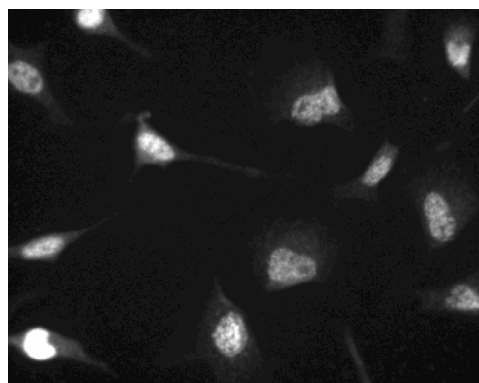
Material Number:	559693
Size:	0.1 mg
Concentration:	0.5 mg/ml
Clone:	DCS-35
Immunogen:	Human Cdk4
Isotype:	Mouse IgG1, κ
Reactivity:	QC Testing: Human Tested in Development: Dog, Rat, Mouse
Target MW:	33 kDa
Storage Buffer:	Aqueous buffered solution containing $\leq 0.09\%$ sodium azide.

Description

Cyclins, cyclin-dependent kinases (Cdks), and cyclin-dependent kinase inhibitors (CdkIs) are essential for cell-cycle control in eukaryotes (reviewed in 1). Cyclins, regulatory subunits, bind to cyclin-dependent kinases (Cdks), catalytic subunits, to form active cyclin-Cdk complexes. Cdk subunits by themselves are inactive and binding to a cyclin is required for their activity. Cyclins A, B1, D and E undergo periodic synthesis and degradation, thereby providing a mechanism to regulate Cdk activity throughout the cell cycle. Additionally, Cdk activity is further regulated by activating and inhibitory phosphorylations, and small proteins (p15, p16, p18, p19, p21 and p27), called CdkIs, that bind to cyclins, Cdks, or cyclin-Cdk complexes. Cdk4 was originally called PSK-J3,2 and following demonstration of its association with D-type cyclins, became known as Cdk4.2 D-type cyclins also associate with Cdks 2 and 5, although Cdk4 appears to be the most abundant partner. The D-type cyclins (D1, D2, and D3) are expressed in response to growth factors or mitogens, and rapidly degrade when mitogens are withdrawn. D cyclins appear to promote G0 to G1 transitions and the rate of G1 progression. For example, cyclin D-Cdk4 and cyclin D-Cdk6 complexes phosphorylate the retinoblastoma protein (Rb) which removes the G1 phase block caused by underphosphorylated Rb. Cdk4 has a molecular weight of ~33 kD.



Western blot analysis of Cdk4. 293 cell lysates were probed with 5 $\mu\text{g/ml}$ (lane 1), 2 $\mu\text{g/ml}$ (lane 2) or 0.5 $\mu\text{g/ml}$ (lane 3) of anti-Cdk4 (clone DCS-35, Cat. No. 68941A). The antibody identifies Cdk4 at ~33 kDa.



Immunofluorescent staining of HeLa (ATCC CCL-2) cells. Cells were seeded in a 96 well imaging plate (Cat. No. 353219) at ~10 000 cells per well. After overnight incubation, cells were stained using the alcohol perm protocol and the anti-Cdk4 antibody. The second step reagent was Alexa Fluor® 555 (Invitrogen). The image was taken on a BD Pathway™ 855 Bioimager system using a 20x objective. This antibody also stained A549 (ATCC CCL-185) and U-2 OS (ATCC HTB-96) cells and worked with both the Triton™ X-100 and alcohol perm protocols (see Recommended Assay Procedure).

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Preparation and Storage

Store undiluted at 4°C.

The monoclonal antibody was purified from tissue culture supernatant or ascites by affinity chromatography.

Application Notes

Application

Western blot	Routinely Tested
Immunoprecipitation	Tested During Development
Bioimaging	Tested During Development
Immunohistochemistry-frozen	Tested During Development

Recommended Assay Procedure:

Bioimaging

- Seed the cells in appropriate culture medium at ~10,000 cells per well in a BD Falcon™ 96-well Imaging Plate (Cat. No. 353219) and culture overnight.
- Remove the culture medium from the wells, and fix the cells by adding 100 µl of BD Cytofix™ Fixation Buffer (Cat. No. 554655) to each well. Incubate for 10 minutes at room temperature (RT).
- Remove the fixative from the wells, and permeabilize the cells using either BD Perm Buffer III, 90% methanol, or Triton™ X-100:
 - Add 100 µl of -20°C 90% methanol or Perm Buffer III (Cat. No. 558050) to each well and incubate for 5 minutes at RT.OR
 - Add 100 µl of 0.1% Triton™ X-100 to each well and incubate for 5 minutes at RT.
- Remove the permeabilization buffer, and wash the wells twice with 100 µl of 1× PBS.
- Remove the PBS, and block the cells by adding 100 µl of BD Pharmingen™ Stain Buffer (FBS) (Cat. No. 554656) to each well. Incubate for 30 minutes at RT.
- Remove the blocking buffer and add 50 µl of the optimally titrated primary antibody (diluted in Stain Buffer) to each well, and incubate for 1 hour at RT.
- Remove the primary antibody, and wash the wells three times with 100 µl of 1× PBS.
- Remove the PBS, and add the second step reagent at its optimally titrated concentration in 50 µl to each well, and incubate in the dark for 1 hour at RT.
- Remove the second step reagent, and wash the wells three times with 100 µl of 1× PBS.
- Remove the PBS, and counter-stain the nuclei by adding 200 µl per well of 2 µg/ml Hoechst 33342 (e.g., Sigma-Aldrich Cat. No. B2261) in 1× PBS to each well at least 15 minutes before imaging.
- View and analyze the cells on an appropriate imaging instrument.

Bioimaging: For more detailed information please refer to http://www.bdbiosciences.com/support/resources/protocols/certified_reagents.jsp

Western blot: For more detailed information please refer to http://www.bdbiosciences.com/pharmingen/protocols/Western_Blotting.shtml

Suggested Companion Products

Catalog Number	Name	Size	Clone
353219	BD Falcon™ 96-well Imaging Plate	NA	(none)
554655	Fixation Buffer	100 ml	(none)
558050	Perm Buffer III	125 ml	(none)
554656	Stain Buffer (FBS)	500 ml	(none)
554002	HRP Goat Anti-Mouse Ig	1.0 ml	(none)

Product Notices

- Since applications vary, each investigator should titrate the reagent to obtain optimal results.
- Please refer to www.bdbiosciences.com/pharmingen/protocols for technical protocols.
- This antibody has been developed and certified for the bioimaging application. However, a routine bioimaging test is not performed on every lot. Researchers are encouraged to titrate the reagent for optimal performance.
- Caution: Sodium azide yields highly toxic hydrazoic acid under acidic conditions. Dilute azide compounds in running water before discarding to avoid accumulation of potentially explosive deposits in plumbing.
- Triton is a trademark of the Dow Chemical Company.

References

Depoortere F, Van Keymeulen A, Lukas J, et al. A requirement for cyclin D3-cyclin-dependent kinase (cdk)-4 assembly in the cyclic adenosine monophosphate-dependent proliferation of thyrocytes. *J Cell Biol.* 1998; 140(6):1427-1439. (Immunogen)
Johnson DG, Walker CL. Cyclins and cell cycle checkpoints. *Annu Rev Pharmacol Toxicol.* 1999; 39:295-312. (Biology)
Matsushima H, Ewen ME, Strom DK, et al. Identification and properties of an atypical catalytic subunit (p34PSK-J3/cdk4) for mammalian D type G1 cyclins. *Cell.* 1992; 71(2):323-334. (Biology)